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## **Comparison between bone–implant OPEN interfaces of microtopographically modifed zirconia and titanium implants**

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**The aim of this study was to investigate the surface characteristics and evaluate the bone–implant interfaces of injection molded zirconia implants with or without surface treatment and compare them with those of conventional titanium implants. Four diferent zirconia and titanium implant groups (n= 14 for each group) were prepared: injection-molded zirconia implants without surface treatment (IM ZrO2); injection-molded zirconia implants with surface treatment via sandblasting (IM ZrO2-S); turned titanium implants (Ti-turned); and titanium implants with surface treatments via sandblasting with large-grit particles and acid-etching (Ti-SLA). Scanning electron microscopy, confocal laser scanning microscopy, and energy dispersive spectroscopy were used to assess the surface characteristics of the implant specimens. Eight rabbits were used, and four implants from each group were placed into the tibiae of each rabbit. Bone-to-implant contact (BIC) and bone area (BA) were measured to evaluate the bone response after 10-day and 28-day healing periods. One-way analysis of variance with Tukey's pairwise comparison was used to fnd any signifcant diferences. The signifcance level was set at α= 0.05. Surface physical analysis showed that Ti-SLA**  had the highest surface roughness, followed by IM ZrO<sub>2</sub>-S, IM ZrO<sub>2</sub>, and Ti-turned. There were no **statistically signifcant diferences (***p* **> 0.05) in BIC and BA among the diferent groups according to the histomorphometric analysis. This study suggests that injection-molded zirconia implants are reliable and predictable alternatives to titanium implants for future clinical applications.**

#### **Abbreviations**



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### m-ZrO<sub>2</sub> Monoclinic zirconia<br>t-ZrO<sub>2</sub> Tetragonal zirconia

Tetragonal zirconia

Implant dentistry has become a predictable and reliable treatment modality for restoring the esthetics and functions of partially and completely edentulous patients. For successful outcomes of implant-supported dental prostheses, osseointegration, the biological fxation of an implant defned as direct bone-to-implant contact  $(BIC)$  without an intervening connective tissue layer, must be achieved<sup>[1,](#page-7-0)[2](#page-7-1)</sup>. Therefore, BIC is a critical indicator for successful osseointegration, which governs the long-term clinical success and survival of implant-supported dental prostheses<sup>[3](#page-7-2)</sup>. For the optimal BIC, the surface characteristics, both physical and chemical, of dental implants play a critical role in the osseointegration process.

Over the last 40 years, both commercially pure titanium (Ti) and titanium alloys have been used for dental implants due to their excellent biocompatibility, mechanical strength, chemical inertness, and high clinical suc-cess rates<sup>[4](#page-7-3)</sup>. Since the 1990s, various microtopographical modification methods for Ti implant surfaces have been introduced by implant manufacturers to accelerate early osseous integration and increase the resistance of the bone–implant interface to functional loading<sup>[5](#page-8-0)</sup>. Among them, roughening Ti implant surfaces at the microscopic level with sandblasting and acid-etching has become very popular due to its high success rate<sup>[6](#page-8-1)</sup>. However, the main disadvantage of Ti implants is their dark grayish color, which may lead to esthetic compromise if buccal bone loss and unfavorable soft-tissue response in gingival biotypes occur<sup>7</sup>.

Therefore, researchers have developed zirconia (zirconium dioxide,  $ZrO<sub>2</sub>$ ) implants in the last 2 decades to mitigate the esthetic disadvantage of titanium implants $^{8,9}$  $^{8,9}$  $^{8,9}$ . The main advantage of  $ZrO_2$  implants is their tooth-like color, which makes them superior to Ti implants in esthetically critical locations, such as the maxillary anterior area. In addition, they possess promising characteristics, such as a high resistance to wear and corrosion, high fracture resistance and fexural strength, biocompatibility, minimal ion-release, and reduced bacterial adhesion and plaque accumulation<sup>[10–](#page-8-5)[12](#page-8-6)</sup>. However, ZrO<sub>2</sub> also has disadvantages, such as a sensitivity to low-temperature degradation (aging), vulnerability to subcritical bending and crack growth, and lower fracture resistance, com-pared to titanium<sup>[13](#page-8-7),14</sup>. It is also challenging to roughen  $ZrO<sub>2</sub>$  implant surfaces because of their hardness<sup>15</sup>. In the literature, various attempts to modify  $ZrO<sub>2</sub>$  surfaces have been reported, such as sandblasting, etching with hydrochloric or hydrofluoric acids<sup>16</sup>, the aggregation of bioactive materials such as hydroxyapatite<sup>17</sup>, plasma spraying<sup>18</sup>, ultraviolet radiation to induce the hydrophilicity of  $ZrO_2^{19}$  $ZrO_2^{19}$  $ZrO_2^{19}$ , and selective infiltration-etching techniques to create a nanoporous surface<sup>20</sup>.

Although both the additive manufacturing and subtractive (milling) manufacturing of  $ZrO<sub>2</sub>$  by computeraided design and computer-aided manufacturing (CAD/CAM) technology have been studied frequently, as reported in the literature<sup>[21](#page-8-15)[–23](#page-8-16)</sup>, the injection molding of  $ZrO<sub>2</sub>$  has been reported in the literature since the 1980s<sup>[24](#page-8-17)</sup>. Tis technique is a type of plasticity-shaping technique that utilizes a binder, plasticizer, lubricant, and coupling agent<sup>[25](#page-8-18)</sup>. The process begins with compounding fine ceramic powders with a blend of polymers or wax in solvent. Then, the organic binders embedded in the molds are removed via thermal pyrolysis or solvent detracting before sintering<sup>26</sup>. Injection molding is suitable for making relatively fine ceramic parts because it offers dimensional reproducibility, requires little or no modification, and produces parts with a sufficient quality for clinical applications<sup>[27](#page-8-20)</sup>. In particular, this technique allows the mass production of ceramic parts at low cost and near-net-shape formation<sup>28</sup>.

Previous animal studies and various case reports have indicated that the osseointegration of  $ZrO<sub>2</sub>$  implants is similar to or even superior to that of Ti implants<sup>[29](#page-8-22)[–31](#page-8-23)</sup>. However, the effects of different surface characteristics (i.e., smooth vs. rough) of  $ZrO<sub>2</sub>$  implants on the osseointegration process are still largely unknown<sup>32</sup>. Furthermore, there is no report on the early bone response around injection-molded ZrO<sub>2</sub> implants compared to that around Ti implants. This in vivo study aimed to compare the bone–implant interfaces of injection-molded  $ZrO<sub>2</sub>$ implants and computer numerical control (CNC)-machined Ti implants. Two types of surfaces were prepared for each material: sandblasted and nonsandblasted surfaces for the injection-molded  $ZrO<sub>2</sub>$  implants and sandblasted, large-grit, acid-etched (SLA) and non-SLA surfaces for the CNC-machined Ti implants. The surface characteristics of the implants were also investigated. The hypothesis underlying this study was that the hardtissue response to the injection-molded  $ZrO<sub>2</sub>$  implants is similar to the osseointegration of Ti implants with SLA surfaces, which are used in dental clinics worldwide.

### **Methods**

**Preparation of implant samples.** A total of 56 screw-shaped dental implant samples (28 ZrO<sub>2</sub> implants (one-piece) and 28 Ti implants) of the same macroscopic shape and dimensions (a diameter of 3.4 mm and a length of 8 mm) were used in this study. Also, ten ZrO<sub>2</sub> discs, which were 15 mm in diameter and 1 mm in thickness, were prepared to find some phase transition of  $ZrO<sub>2</sub>$  after surface modification. The  $ZrO<sub>2</sub>$  implants were manufactured using an injection molding technique (Vatech Acucera, Seoul, Korea). Tis process involves mixing zirconia powders with modifiers and shaping a uniform and homogeneous mixture into a mold. The Ti implants were manufactured by a computer numerical controlled (CNC) milling technique (Deep Implant System, Inc., Seongnam, Korea). Disc-shaped green compacts were prepared by cold isostatic press of powder mixtures and then sintered. Surface modification with sandblasting was performed on half of the  $ZrO<sub>2</sub>$  implants and the ZrO<sub>2</sub> discs, while half of the Ti implant samples were surface-modified with SLA treatment, i.e., sandblasted with large-grit alumina  $(A<sub>1</sub>, O<sub>3</sub>)$  particles and etched with hydrochloric acid (SLA surface; Deep Implant System, Inc., Seongnam, Korea). After sandblasting, the  $ZrO<sub>2</sub>$  implants (14 implants) and discs (5 discs) were treated by hot isostatic pressing (HIP) at 1380 °C and 138 MPa. Then, the samples were divided into four experimental groups:

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- (1) Group 1 = IM ZrO<sub>2</sub> (injection-molded ZrO<sub>2</sub> implants or ZrO<sub>2</sub> discs without sandblasting modification)<br>(2) Group 2 = IM ZrO<sub>2</sub>-S (injection-molded ZrO<sub>2</sub> implants or ZrO<sub>2</sub> discs with sandblasting modification)
- (2) Group 2=IM  $ZrO_2-S$  (injection-molded  $ZrO_2$  implants or  $ZrO_2$  discs with sandblasting modification)<br>(3) Group 3=Ti-turned (Ti implants without SLA modification as a negative control)
- Group 3 = Ti-turned (Ti implants without SLA modification as a negative control)
- (4) Group 4=Ti-SLA (Ti implants with SLA modifcation as a positive control)

Assessment of surface characteristics. The implant sample surfaces were photographed by field emission-scanning electron microscopy (FE-SEM; S-4700, Hitachi, Tokyo, Japan). Surface parameters for the sample topography were measured by confocal laser scanning microscopy (CLSM; LSM 800, Carl Zeiss AG, Oberkochen, Germany). The acquired images were analyzed using ConfoMap software, and specific areas of interest were selected. Subsequently, the surface topography was quantifed in terms of Sa (arithmetical mean height of a surface), the absolute value of the diference in height of each point compared to the arithmetical mean of the surface, and Sdr (developed interfacial area ratio), the proportion of the additional surface area contributed by the texture within the defned planar area. Each sample was analyzed at 3 selected sites (the upper, middle and lower fank), the values of which were averaged, and the average value was assigned as the representative value for the sample<sup>[22](#page-8-25),[33](#page-8-26)</sup>. The topography was measured in terms of Sa values (arithmetical mean height) and Sdr values (developed interfacial area ratio). In addition, the chemical composition of each sample was analyzed with an energy-dispersive spectroscopy (EDS) device (EMAX, Horiba, High Wycombe, United Kingdom). For evaluating the phase transition of  $ZrO<sub>2</sub>$ , the  $ZrO<sub>2</sub>$  disc surfaces were analyzed using a high resolution X-ray difractometer (SmartLab, Rigaku, Tokyo, Japan) with Cu Kα radiation (wavelength=1.54 Å) and 45 kV/200 mA. To quantify the molar fraction of the content of monoclinic  $ZrO<sub>2</sub>(X<sub>m</sub>)$ , the following equations were employed:

$$
X_m = \frac{I_m(111) + I_m(\bar{1}11)}{I_m(111) + I_m(\bar{1}11) + I_t(101)}
$$

where  $I_t$  and  $I_m$  represent the intensity of the tetragonal (101) and monoclinic (111) and  $(\bar{1}11)$  peaks<sup>[34](#page-8-27),[35](#page-8-28)</sup>. The peak intensity was obtained using MDI Jade 6 sofware (Materials Data Inc., Livermore, CA, USA).

**In vivo surgery.** Eight male New Zealand white rabbits (age: 3–4 months old; weight: 2.5–3.0 kg) were used in this in vivo study, which was approved by the Institutional Animal Research Ethics Committee of Cronex (CRONEX-IACUC: 202108011, Hwaseong, Korea) and conducted according to the Animal Research: Reporting In Vivo Experiments (ARRIVE) guidelines<sup>36</sup>. All methods were performed in accordance with the relevant guidelines and regulations. The experimental animals acclimatized in separate cages for two weeks before surgery. Each rabbit received four implant samples; two implants were placed in each tibia bone of the hind legs. The  $ZrO<sub>2</sub>$  and Ti implants were inserted based on the split-plot design (Fig. [1\)](#page-3-0). For anesthesia, a combination of 15 mg/kg tiletamine hydrochloride and zolazepam hydrochloride (Zoletil 50; Virbac Korea Co., Ltd., Seoul, Korea) and 5 mg/kg xylazine (Rompun; Bayer Korea, Ltd., Seoul, Korea) was administered intramuscularly. Then, the hind legs were shaved and disinfected with an antiseptic surgical scrub of 7.5% povidone-iodine (Betadine; Korea Pharma, Seoul, Korea). Afer site preparation, local infltration anesthesia of 2% lidocaine hydrochloride with 1:100,000 epinephrine (Yuhan Company, Seoul, Korea) was administered at the surgical sites. The tibial bones were exposed by full-thickness incisions from the skin to the periosteum. The surgical sites on the tibiae bones were prepared with rotating implant drills and engines under copious irrigation with sterile saline solution. The final drill size was 3.0 mm, and 32 sample implants from experimental groups 1, 2, 3 and 4 were placed with primary stability ( $\geq$  20 Ncm) using a torque wrench, according to the manufacturer's instructions. Afer the implant placement surgeries, the muscle and fascia were sutured with resorbable 4–0 Vicryl sutures (Coated Vicryl; Ethicon, Raritan, NJ, United States), and the outer dermis was closed with nylon (Blue nylon; Ailee, Busan, Korea). All rabbit specimens were housed in individual cages and administered the postoperative antibiotic prophylaxis of enrofoxacin (Biotril, Komipharm International, Siheung, Korea).

**Assessment of histology and histomorphometry.** Four rabbits were sacrificed 10 days after the implant placement (Rabbit 1–4), and the remaining 4 rabbits were sacrifced 28 days afer the implant placement (Rabbit 5–8) by an overdose of potassium chloride intravenously under anesthesia for histologic assessment. The bones and connective tissues that surrounded the implant samples were surgically harvested en bloc. These blocks were fxed in 10% neutral bufered formalin for two weeks and then dehydrated with ethanol, followed by embedding in light-curing resin (Technovit 7200 VLC, Kulzer, Wehrheim, Germany). A series of cutting and grinding devices (EXAKT system; EXAKT Apparatebau, Norderstedt, Germany) was used to cut and grind the embedded blocks into slides with a thickness of less than approximately 50  $\mu$ m<sup>22,37</sup>. The slides were stained with modified Goldner's Masson trichrome staining solution for examination under a light microscope. This staining technique allows for easy discrimination between newly formed bone (stained red) and exiting mature bone (stained blue)<sup>[38](#page-8-31)</sup>. The bone-to-implant interfaces were measured by the degree of BIC and bone area (BA) at the best three consecutive threads $22,39$  $22,39$ . The histologic evaluation was performed using a light microscope (DM2700M, Leica Microsystems CMS GmbH, Wetzlar, Germany) and an attached digital camera (DMC5400, Leica Microsystems CMS GmbH, Wetzlar, Germany). An image analysis system (ImageJ 1.60, NIH, Bethesda, MD, United States) was used to analyze the acquired images.

**Statistical analysis.** Most of the outcome variables for data normalization were accepted when the Shapiro–Wilk test was used ( $p > 0.05$ ). Descriptive statistics are shown as the means and standard deviations (SDs). One-way analysis of variance (ANOVA) was used to analyze diferences in the mean values of the surface param-



<span id="page-3-0"></span>**Figure 1.** Four implant groups used in this study. 1: injection-molded ZrO<sub>2</sub> implant; 2: sandblasted injectionmolded  $ZrO<sub>2</sub>$  implant; 3: turned Ti implant; 4: SLA Ti implant. This schematic diagram shows that these implants were placed in rabbit tibiae according to the split-plot design. The saw-edged circles represent the implants whose surfaces were microtopographically modifed, while the smooth edged ones represent implants with no microtopographical modification. Black represents Ti, and white represents  $ZrO_2$ . R=right tibia; L=left tibia.

eters and histomorphometric data of the diferent types of implants in this study. When a signifcant diference was found, Tukey's honestly signifcant diference (HSD) was further applied for a pairwise comparison. In addition, the independent t test was used to compare the BIC and BA values of the 2 healing periods (10 days vs. 28 days). Statistical sofware, R version 4.1.0 (R Foundation for Statistical Computing, Vienna, Austria), was used for all statistical evaluations. The statistical significance level was set at  $\alpha$  = 0.05.

**Ethics approval and consent to participate.** Ethical approval was sought from the Institutional Animal Care and Use Committee of CRONEX Co., Ltd., Hwaseong, Korea, and the animal experiment was conducted in accordance with the Animal Reporting in Vivo Experiments (ARRIVE) guidelines (202108011).

**Consent for publication.** All authors are aware of the publication of this work.

### **Results**

**Surface characteristics.** The FE-SEM images of the implant surfaces of the 4 different experimental groups are shown in Fig. [2](#page-4-0)A. The IM ZrO<sub>2</sub> (Group 1) implants showed microcracks, porosities, and grain structures that are typically observed for sintered  $ZrO<sub>2</sub>$ . In contrast, the IM  $ZrO<sub>2</sub>$ -S (Group 2) implants showed very rough surfaces with slate-like profiles and lower porosities. The Ti-turned (Group 3: negative control) implants showed smooth and fat surfaces with continuous straight lines that ran in one direction. However, the Ti-SLA (Group 4: positive control) implants exhibited a rough, irregular, porous, and honeycomb-like appearance.

The surface roughness parameters of the experimental groups were measured in terms of Sa and Sdr values (Fig. [2](#page-4-0)B,C). Based on the mean Sa values and SDs, the Ti-SLA implants had the highest Sa value (1.68  $\mu$ m ± 0.07  $\mu$ m), followed by the IM ZrO<sub>2</sub>-S implants (1.10  $\mu$ m ± 0.13  $\mu$ m), IM ZrO<sub>2</sub> implants  $(0.64 \mu m \pm 0.12 \mu m)$ , and Ti-turned implants  $(0.52 \mu m \pm 0.21 \mu m)$ . There were statistically significant differences among the Sa values of all groups, except between the IM ZrO<sub>2</sub> and Ti-turned groups. The mean Sdr values of the Ti-SLA implants (238.78% $\pm$ 3.03%) were significantly higher than those of the other three groups. However, there were no significant differences among the Sdr values of these three groups: IM ZrO<sub>2</sub> (49.04% $\pm$ 31.68%), IM  $ZrO<sub>2</sub>$ -S (78.77% ± 50.02%), and Ti-turned (123.66% ± 37.54%).

The EDS findings are shown in Table [1](#page-5-0). Titanium (Ti), carbon (C), and oxygen (O) were detected in the Ti implants, while zirconium  $(Zr)$ , carbon  $(C)$ , and oxygen  $(O)$  were detected in the  $ZrO<sub>2</sub>$  implants. The X-ray diffraction patterns of the  $ZrO_2$  discs are shown in Fig. [3](#page-5-1).  $ZrO_2$  discs without sandblasting modification only presented the tetragonal ZrO<sub>2</sub> peak. ZrO<sub>2</sub> discs with sandblasting modification presented the (111) and ( $\bar{1}11\bar{)}$ peaks for the monoclinic phase (m-ZrO<sub>2</sub>) and the (101) peak for the tetragonal phase (t-ZrO<sub>2</sub>). However, it was

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<span id="page-4-0"></span>**Figure 2.** Surface characteristics of the ZrO<sub>2</sub> and Ti specimens investigated in this study. (a) FE-SEM images of the surfaces of IM ZrO<sub>2</sub> (injection-molded zirconia without surface treatment), IM ZrO<sub>2</sub>-S (injection-molded zirconia implant with sandblasting surface treatment), Ti-turned (turned titanium implant), and Ti-SLA (SLA titanium implant) at 2000×(lef) and 5000×(right) magnifcations. Scale bars=10 μm. (**b**) CLSM images of surface roughness of implant surfaces (3D views of color-coded height maps with texture overlay). (**c**) Surface roughness parameters (Sa and Sdr) of the implant surfaces. Data are presented as the means  $\pm$  SDs (n=3). \**p*<0.05 and \*\**p*<0.01.

	Atomic percentage (%)				
Group $(n=3)$				Zr	Ti
IM $ZrO2$	56.67	07.47	0.00	35.85	00.00
IM $ZrO2-S$	48.58	24.56	0.00	26.85	00.00
Ti-turned	05.56	11.86	0.00	00.00	82.54
Ti-SLA	04.04	14.24	0.00	00.00	81.72

<span id="page-5-0"></span>**Table 1.** Element composition analysis of the  $ZrO<sub>2</sub>$  and Ti implant surfaces according to energy dispersive spectroscopy. *C* carbon, *O* oxygen, *N* nitrogen, *Zr* zirconium, *Ti* titanium.



<span id="page-5-1"></span>**Figure 3.** X-ray diffraction (XRD) patterns of the ZrO<sub>2</sub> discs (**a**) in the range of 20–80° 2θ are presented. More detailed XRD patterns are also shown in the range of 27-33° 2 $\theta$  (b). Compared to the XRD peaks of the ZrO<sub>2</sub> discs before sandblasting (A), the presence of the monoclinic phase was detected after the  $ZrO<sub>2</sub>$  discs were sandblasted (B). The monoclinic phase disappeared upon hot isostatic pressing of the sandblasted ZrO<sub>2</sub> discs (C).

clearly seen that while the  $ZrO<sub>2</sub>$  discs with sandblasting modification presented both the peaks corresponding to the tetragonal and monoclinic phases, only the peak of tetragonal phase was detected afer HIP. As seen in Table [2](#page-5-2), the rate of the tetragonal-to-monoclinic phase transformation increased with sandblasting modifcation. It was found that the amount of m-ZrO<sub>2</sub> was 40.32% of the total  $ZrO<sub>2</sub>$  after sandblasting modification. After HIP, this amount of monoclinic phase was totally transformed into  $t$ - $ZrO<sub>2</sub>$ .

**Analysis of histology and histomorphometry.** All the implant samples successfully osseointegrated after 10 days of healing and 28 days of healing (Supplementary Fig. S1). In the histological analysis of the  $ZrO<sub>2</sub>$ and Ti implants stained with modifed Goldner's Masson trichrome staining solution, new bone formation was found along the bone–implant interfaces. In the cortical bone area of the tibiae, the exiting mature bones were stained blue, while newly formed immature bones were stained red and detected at the implant threads and around the mature bone (Fig. [4\)](#page-6-0).

The mean BIC and BA values and SDs of the  $ZrO<sub>2</sub>$  and Ti implants at the 10-day and 28-day marks are shown in Fig. [5](#page-7-4) (Supplementary Tables S1, S2). Although the BICs (%) of the IM  $ZrO<sub>2</sub>$ -S and Ti-SLA implants were higher than those of the Ti-turned and IM ZrO<sub>2</sub> implants at the 10-day mark, the differences were not statistically signifcant. At the 28-day mark, the BIC (%) of the Ti-SLA implants was signifcantly higher than that of the Ti-turned implants ( $p=0.04$ ). However, the other groups showed no significant differences. When the mean values and SDs of BA (%) were calculated, no statistically significant differences were found among the  $ZrO<sub>2</sub>$ and Ti implants for both healing periods  $(p > 0.05)$  (Fig. [5\)](#page-7-4).



<span id="page-5-2"></span>**Table 2.** The tetragonal-to-monoclinic phase transformation in ZrO<sub>2</sub> discs with and without sandblasting modification and after HIP. \*Monoclinic  $ZrO<sub>2</sub>$ . †Hot isostatic pressing.



<span id="page-6-0"></span>**Figure 4.** Histological views and histomorphometric data for bone responses to the ZrO<sub>2</sub> and Ti implants. Light microscopic views of the implants with modifed Goldner's Masson trichrome staining at 10 and 28 days after installation into rabbit tibiae (scale bars from left to right: 1 mm at ×12.5, 200  $\mu$ m at ×50, and 100  $\mu$ m at × 100 magnification). The exiting mature bone was stained blue (white arrow), and new immature bone or osteoid was stained red (white-edged arrowhead) at the bone–implant interfaces.

### **Discussion**

The roughness parameters (Sa and Sdr) of the surface-treated implants (Groups 2 and 4) were significantly higher than those of the ZrO<sub>2</sub> and Ti implant groups without surface treatment (Groups 1 and 3). The authors also found that the BIC% and BA% of the  $\bar{\text{IM ZrO}}_2$  implants were not significantly different from those of the Ti implants (both Ti-turned and Ti-SLA) after 10-day and 28-day healing periods. These results are in agreement with the findings of previous studies that compared SLA-treated IM  $ZrO<sub>2</sub>$  and Ti implants in mini pig maxillae models<sup>40</sup>, canine models<sup>[41](#page-8-34)</sup>, and rabbit tibia models<sup>[42](#page-8-35)</sup>. Another significant finding was that the surface-treated  $ZrO<sub>2</sub>$  implants showed enhanced bone integration at the implant surface compared to the untreated ZrO<sub>2</sub> implants. This finding is in line with the results of other studies by Mihatovic in 2017 and Schünemann in 2019<sup>43,44</sup>. In addition, the Ti-SLA implants demonstrated a signifcantly higher bone response than the Ti-turned implants. Tis fnding is also comparable to findings in other studies<sup>[45,](#page-8-38)46</sup>. It can be deduced that the rough surface microtopography of

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<span id="page-7-4"></span>

ZrO2 implants generated by the proper surface modifcation can infuence early bone response and long-term sustainability.

In this study, the authors used 10-day and 28-day healing periods for bone formation in a rabbit tibia model based on previous studies<sup>[47](#page-8-40),[48](#page-9-0)</sup>. These healing periods of rabbit specimens are equivalent to 1-month and 3-month healing periods of humans, according to Roberts, who demonstrated that the bone healing of rabbits was approximately three times faster than that of humans<sup>49</sup>. Although rabbits are regarded as commonly used and well-established animal models for investigating the osseointegration process, some disadvantages include site limitations and mismatched microstructures when compared to those of human bone. In addition, histomorphometric analysis has some disadvantages, although BIC and BA have become the most popular parameters. For example, they are only measured in 2 dimensions without consideration of the entire implant, and they may be afected by the quality and quantity of the surrounding bones<sup>50</sup>. There were slight discrepancies in the geometry of the  $ZrO<sub>2</sub>$ and Ti implants that might have affected the tissue reaction at the bone–implant interface. The small sample size employed in this study also presents a limitation. Te absence of signifcant diferences observed in the in vivo experiments could be attributed to this small sample size. Further studies are needed to develop methods for the predictable early bone response and long-term osseointegration of IM ZrO<sub>2</sub> implants with a double-blinded study design and a larger sample size. Furthermore, it is needed to investigate the optimal surface roughness of  $ZrO<sub>2</sub>$  implants for osseointegration in subsequent studies.

#### **Conclusion**

Based on the findings of this study, it is evident that appropriate surface treatment of  $ZrO<sub>2</sub>$  implants is essential for promoting early peri-implant bone formation. Considering the recognized advantages of simplicity, mass production capability, and economic feasibility associated with the injection molding technique, utilizing injection molded zirconia dental implants, coupled with suitable surface modifcations, could present a promising alternative to conventionally manufactured titanium dental implants for future clinical applications.

#### **Data availability**

All data generated or analyzed during this study are included in this published article.

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#### **Author contributions**

I.L.Y. conceptualized this study. M.K.T., Y.S.K., J.M.K. and Y.H.J. performed the experiments. M.K.T., J.M.K. and I.L.Y. analyzed and interpreted the data. M.K.T., Y.S.K. and J.M.K. were major contributors in writing the manuscript. Y.S.K., Y.H.J., J.S.H. and I.L.Y. reviewed and edited the manuscript. M.K.T., Y.S.K. and J.M.K. contributed equally to this work. All authors read and approved the fnal manuscript.

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#### **Competing interests**

The authors declare no competing interests.

#### **Additional information**

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